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Synaptic protein CSF levels relate to memory scores in individuals without dementia

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Abstract

Background We investigated how cerebrospinal fluid levels of synaptic proteins associate with memory function in normal cognition (CN) and mild cognitive impairment (MCI), and investigated the effect of amyloid positivity on these associations.

Methods We included 242 CN (105(43%) abnormal amyloid), and 278 MCI individuals (183(66%) abnormal amyloid) from the European Medical Information Framework for Alzheimer's Disease Multimodal Biomarker Discovery (EMIF-AD MBD) and the Alzheimer's Disease Neuroimaging Initiative (ADNI). For 181 (EMIF-AD MBD) and 36 (ADNI) proteins with a synaptic annotation in SynGO, associations with word learning recall were analysed with linear models.

Results Subsets of synaptic proteins showed lower levels with worse recall in preclinical AD (EMIF-AD MBD: 7, ADNI: 5 proteins, none overlapping), prodromal AD (EMIF-AD MBD only, 27 proteins) and non-AD MCI (EMIF-AD MBD: 1, ADNI: 7 proteins). The majority of these associations were specific to these clinical groups.

Conclusions Synaptic disturbance-related memory impairment occurred very early in AD, indicating it may be relevant to develop therapies targeting the synapse early in the disease.

Keywords Synaptic proteins, Cerebrospinal fluid proteomics, Memory performance, Early Alzheimer's disease

Background

Memory impairment is a key feature of Alzheimer's disease (AD) [13, 14]. Decline in memory function precedes dementia onset, starting already in the preclinical stage of AD [15]. A better understanding of mechanisms underlying memory loss can help the development of novel therapies.

Previous studies have shown that memory scores are associated with reduced synaptic density [27, 28, 35–40] but this research was mostly conducted post-mortem in AD patients who were in late disease stages. Synaptic protein levels can also be assessed in vivo in cerebrospinal fluid (CSF). So far, studies have focused on few or

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Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf.

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single synaptic markers in CSF. For example, neurogranin (NRGN) is a widely studied postsynaptic protein, showing higher levels with worse memory performance in prodromal AD [12], although contradicting results have been found for memory associations in cognitively normal (CN) individuals [9, 12, 19] and in AD dementia [29, 41]. Associations of synaptosomal-associated protein 25 (SNAP25) and neuromodulin (also called growth-associated protein 43, GAP43) with memory functioning depended on disease stage, with associations in early but not in later AD [31]. Many additional synaptic proteins exist, for which it is largely unknown if they correlate with memory functioning in the early stages of AD before dementia onset. Furthermore, individuals can have mild cognitive impairment (MCI) with *normal* amyloid (i.e., non-AD MCI), and it remains unclear what mechanisms may explain impaired memory in such individuals.

Using a proteomics approach of CSF, we aimed to analyse which synaptic proteins are related to memory scores in older non-demented individuals. We hypothesized that synaptic protein levels in CSF would be associated with worse memory scores, and that a subset of these associations would be specific for AD pathology and/or depend on cognitive stage. In two independent cohorts, we analysed cross-sectional associations of synaptic protein levels in CSF with memory scores in preclinical AD and prodromal AD, and in CN amyloid normal controls and non-AD MCI, to test if memory associations were specific for AD pathology.

Methods

Study participants

Two independent study cohorts, the European Medical Information Network Alzheimer's disease multi-modal biomarker discovery study (EMIF-AD MBD) [5] and Alzheimer's Disease Neuroimaging Initiative (ADNI) [1] were used in these analyses. All participants provided informed consent to participate in these studies. We selected individuals who were CN and individuals with a clinical diagnosis of MCI if they had memory test scores and CSF proteomic data available. In EMIF-AD MBD, CN was defined in all centres based on neuropsychological examination scores within 1.5 standard deviation (SD) of age-, sex- and education adjusted standards, and in four centres with additional criteria (see [5] for more details). MCI was defined using cohort-specific criteria (in all cohorts Petersen criteria [32] except for Lausanne, where Winblad criteria [48] were used and Clinical Dementia Rating (CDR) of 0.5).

The ADNI (adni.loni.usc.edu) was launched in 2003 as a public-private partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic resonance

imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment (MCI) and early Alzheimer's disease (AD). For up-to-date information, see www.adni-info.org. Detailed information about the definitions of CN and MCI in ADNI, and methodology of the memory testing can be found in the general procedures manual of ADNI [1]. In short, CN was defined as absence of memory complaints, normal scores on the Logical Memory II subscale (delayed Paragraph Recall) from the Wechsler Memory Scaled – Revised, Mini-Mental State Examination (MMSE) score ≥ 24 and CDR of 0. MCI was defined by a memory complaint, abnormal scores on the Logical Memory II subscale, MMSE ≥ 24 and CDR of 0.5.

For the analyses, we characterized participants from EMIF-AD MBD and ADNI into four diagnostic groups depending on their clinical and amyloid status: CN individuals with normal amyloid (controls), CN individuals with abnormal amyloid (preclinical AD), MCI individuals with normal amyloid (non-AD MCI) and MCI individuals with abnormal amyloid (prodromal AD).

CSF amyloid, t-tau and p-tau biomarkers

For EMIF-AD MBD, we used cut-points for amyloid status that were previously determined with gaussian mixture modelling for each center [44]. T-tau and p-tau were measured in local laboratories using either Innostest (Fujirebio Europe, Gent, Belgium) (4 cohorts), or with INNO-BIA AlzBio3 (Fujirebio Europe), and we used local cut-offs to determine abnormality [5]. In ADNI, CSF amyloid, t-tau and p-tau were measured using multiplex xMAP Luminex platform (Luminex Corp, Austin, TX) with INNO-BIA AlzBio3 (Fujirebio Europe), with abnormality defined as amyloid < 192 pg/ml, t-tau > 93 pg/ml and p-tau > 23 pg/ml.

Proteomics

In EMIF-AD MBD, proteomics was performed with the tandem mass tag (TMT) technique using 10+1 plexing [5] as previously described [3, 25, 42]. In addition, in a central laboratory (Gothenburg University, Sweden), levels of neurofilament light (NEFL) were determined with the NF-Light assay (UmanDiagnostics, Umeå, Sweden) and NRGN with an in-house immunoassay [34]. All protein levels were natural log-transformed. This dataset included 2537 proteins in 306 individuals (missing values; mean 6%, range 0 to 99%).

In ADNI, two proteomic datasets were used. In the first dataset, proteomic data were generated using LC/MS-MRM (panel developed by Caprion Proteome Inc.) ("MRM Assays: Caprion," n.d.) and the Rules-Based

Medicine (RBM) (“Myriad RBM Operational Procedures White Paper—Myriad RBM,” n.d.) platform. These included 202 proteins in 214 individuals (missing values; mean 0.3%, range 0 to 45%). We used the pre-processed and quality checked normalized data publicly available on the ADNI website (adni.loni.usc.edu). We natural log-transformed and Z-scored individual RBM measurements and MRM peptide measurements. Next, we generated protein scores by averaging MRM peptides and correlating RBM proteins that were mapped to the same protein (r at least 0.5). Protein levels in both datasets were standardized relative to the mean and standard deviations from controls with normal amyloid, t-tau and p-tau.

Recently, another mass spectrometry proteomics analysis was performed in ADNI (CSF 48 panel) and provided data on 48 proteins in 706 individuals [11]. Proteomic data from this panel was used as the second dataset in this study. Our main analyses, however,

were performed in the Caprion/RBM dataset as these included the largest number of proteins and the second dataset served for validation purposes.

Synapse proteins

We used Synaptic Gene Ontologies to identify synaptic proteins (SynGO) [18], “SynGO—Synaptic Gene Ontologies and annotations,” n.d.), a curated database with detailed annotation of known synapse-related proteins. An additional criterion for inclusion of proteins in our analyses was that proteins were measured in at least 25 individuals per diagnostic subgroup. In ADNI Caprion/RBM and CSF 48 panel, respectively 36 synaptic proteins (17% of 202 total) and 10 synaptic proteins (21% of 48 total) were identified. In EMIF-AD MBD 181 synaptic proteins (13% out of 1350 total) were identified (Fig. 1).

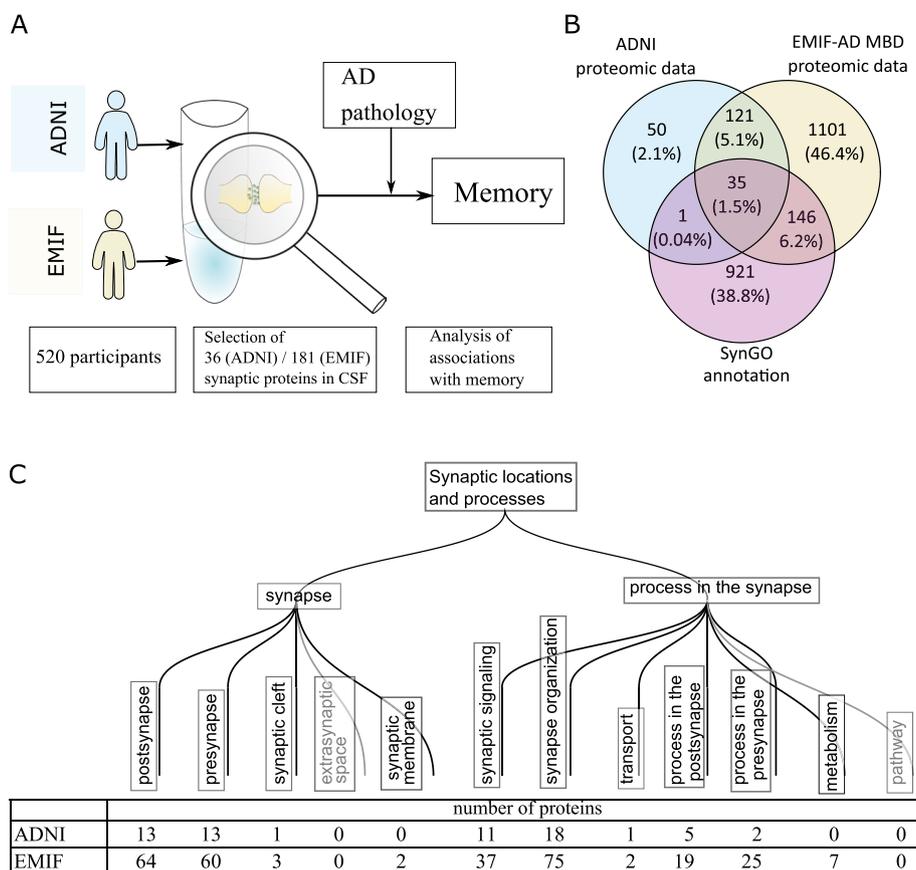


Fig. 1 Characterization of synaptic proteomic dataset. **A** Schematic overview of study design. **B** Overlap of ADNI proteomics and EMIF-AD MBD proteomics with SynGO annotation used to select synaptic proteins. **C** SynGO locations and processes for which synaptic proteins were included in the datasets. One protein can have multiple annotations for different locations and processes. The SynGO annotation uses a hierarchical organisation, and we show here the first and second level of subprocesses, with proteins in lower subprocesses categorized with the respective second level subprocess

Memory tests

We measured memory functioning with both immediate and delayed recall memory tests, because immediate and delayed recall may differ in their sensitivity to detect decline in different stages of AD [4, 16, 45]. In EMIF-AD MBD, memory function was assessed with cohort specific tests (auditory verbal learning test (AVLT): 3 cohorts; Consortium to Establish a Registry for Alzheimer's Disease wordlist (CERAD), Free and Cued Selective Reminding Test (FCSRT) and RI-48 in the remaining 3 cohorts). Test scores were Z transformed according to age, sex and education norms (as detailed in [5]), and values ranged between -5 to 5.

In ADNI, we used the Rey auditory verbal learning test (RAVLT, immediate recall: sum of correctly recalled words over 5 trials, range 0–75; delayed recall: correctly recalled words after 30-min delay, range 0–15). We transformed the memory scores into age-, education- and sex-adjusted Z-scores, using adjustment factors for age, education and sex that were estimated from linear models for memory performance of all ADNI controls.

Statistical analyses

Differences in demographic variables between groups were assessed with chi-square test, Mann–Whitney U test, Student's t-test or Kruskal–Wallis test when appropriate. Unless indicated otherwise, effects were considered significant at a p -value < 0.05. We first tested if protein levels differed depending on diagnostic group with Analysis of variance (ANOVA), and if so, performed two-sided t-tests between the diagnostic groups. To account for the multiple comparisons (in total 6 tests), we considered differences between the diagnostic groups significant at a false discovery rate (FDR)-adjusted p -value < 0.05. Next, we tested associations between memory test scores (outcome) and synaptic protein level (predictor) with linear models that included the covariates diagnostic group, age, sex and years of education (model 1) and additionally the interaction between protein level and diagnostic group (model 2). To report the effects of protein levels on memory scores across diagnostic groups, we selected proteins with a diagnostic group-interaction p -value \geq 0.1. To report the effects of protein levels on memory scores that depended on diagnostic group, we selected proteins if they had a significant interaction with diagnostic group (interaction p -value < 0.1), and calculated associations with memory scores separately in the different diagnostic groups. For reference, we also provide statistical details of both models for all proteins (Supplementary Table 1). We then tested if the memory-associated proteins in our analyses were enriched for specific synaptic functions with the SynGO website ("SynGO—Synaptic Gene Ontologies

and annotations," n.d.), and selected biological process ontology terms for the enrichment analyses when they included at least 3 genes. For visualisation purposes, we calculated composite scores of synaptic proteins, by averaging proteins which showed an interaction (interaction p -value < 0.1) with diagnostic group on immediate or delayed recall scores. All analyses were run in R 4.1.2 "Bird Hippie". *Emmeans* 1.4.2 was used for estimation of regression coefficients.

Results

We included 137 controls, 105 individuals with preclinical AD, 183 individuals with prodromal AD and 95 individuals with non-AD MCI (Table 1). In both studies, the preclinical and prodromal AD groups had more Apolipoprotein E (*APOE*) ϵ 4 carriers and tended to have higher CSF t-tau and p-tau levels than controls. Compared to EMIF-AD MBD participants, ADNI participants were older (all groups), had received longer education (all groups) and had lower memory test scores (controls, preclinical and prodromal AD) (Table 1, Supplementary Fig. 1).

We selected 181 synaptic proteins in EMIF-AD MBD and 36 synaptic proteins in ADNI that were present in at least 25 individuals for each diagnostic group (Fig. 1). In EMIF-AD MBD, 30 of these proteins differed between diagnostic groups (ANOVA p -value < 0.05; supplementary Table 2). Relative to controls, individuals with preclinical AD showed lower levels of VGF, NPTX2, BDNF, CDH2 and higher levels of FXYP6. Compared to controls, individuals with prodromal AD showed lower levels of NPTX2 and higher levels of 9 proteins (YWHAE, YWHAH, YWHAZ, NEFL, NRG1, GAP43, TPD52, VAPA and AKR1A1). The levels of these proteins were typically also higher in prodromal AD compared to preclinical AD and non-AD MCI (Fig. 2A, Supplementary Table 2, Supplementary Table 3). There was no specific association with synaptic locations (pre- and post-synapse) or synaptic functions (synaptic signalling or organization). In ADNI Caprion/RBM, only four proteins showed different levels between diagnostic subgroups (Fig. 2A, Supplementary Table 2). As in EMIF-AD MBD, NRG1 and NEFL were increased in prodromal AD compared to controls, and were also increased compared to individuals with preclinical AD or non-AD MCI (Fig. 2B). FGA and PLG were increased in non-AD MCI relative to prodromal AD and/or controls.

In ADNI Caprion/RBM, we found that several proteins showed an opposite pattern or couldn't be reproduced as it did not fit the inclusion criteria (i.e. measured in > 25 individuals). For example, VGF was decreased in AD in the EMIF-AD cohort while it was increased in ADNI. In addition, 14–3–3 proteins such as YWHAZ were

Table 1 Demographics of study participants in EMIF-AD MBD and ADNI

	EMIF-AD MBD				ADNI			
	Controls	Preclinical AD	MCI with normal amyloid	Prodromal AD	Controls	Preclinical AD	MCI with normal amyloid	Prodromal AD
n	87	73	66	80	50	32	29	103
Age in years, mean ± sd	65.9 ± 7.8 ^{ad}	66.8 ± 8.3 ^{af}	67.2 ± 8 ^{ag}	70.3 ± 6.7 ^{adfg}	75.3 ± 5.5 ^a	76.1 ± 5.6 ^a	75.3 ± 7.8 ^a	74.7 ± 7.2 ^a
Sex, female (%)	43(49%)	39(53%)	29(44%) ^a	45(56%) ^a	25(50%) ^c	15(47%) ^e	5(17%) ^{ace}	38(37%) ^a
APOE-e4 carrier-ship, 1–2 alleles (%)	22(25%) ^{bd}	44(60%) ^{be}	21(32%) ^{aeg}	55(69%) ^{dg}	5(10%) ^{bd}	16(50%) ^{be}	3(10%) ^{aeg}	67(65%) ^{dg}
MMSE score, mean ± sd	28.9 ± 1.2 ^{cd}	28.7 ± 1.2 ^{aef}	26.6 ± 2.6 ^{ce}	26.7 ± 2.5 ^{df}	28.9 ± 1 ^{cd}	29.2 ± 1 ^{aef}	27.4 ± 1.8 ^{ce}	26.8 ± 1.7 ^{df}
Delayed recall Z-score, mean ± sd	0.444 ± 0.99 ^{acd}	0.337 ± 0.95 ^{aef}	−1.28 ± 1.1 ^{ce}	−1.23 ± 1.2 ^{adf}	0.0053 ± 1 ^{acd}	−0.254 ± 1.2 ^{aef}	−1.52 ± 1 ^{ceg}	−1.99 ± 1.1 ^{adfg}
Immediate recall Z-score, mean ± sd	0.523 ± 0.93 ^{acd}	0.252 ± 0.97 ^{ef}	−1.33 ± 1.4 ^{ce}	−0.952 ± 1.3 ^{adf}	0.128 ± 0.96 ^{acd}	0.158 ± 1.1 ^{ef}	−0.985 ± 1.1 ^{ce}	−1.43 ± 1.2 ^{adf}
Education in years, mean ± sd	12.7 ± 3.4 ^{acd}	12.6 ± 3.6 ^{aef}	10.5 ± 3.6 ^{ace}	11.1 ± 3.4 ^{adf}	15.6 ± 2.8 ^a	15.7 ± 3.4 ^a	16.4 ± 2.8 ^a	15.8 ± 3 ^a
CSF amyloid, mean ± sd	0 ± 1 ^{bcd}	−1.42 ± 0.57 ^{abe}	0.425 ± 1.2 ^{ceg}	−1.26 ± 0.86 ^{adg}	0 ± 1 ^{bd}	−4.35 ± 1.6a ^{bef}	−0.0274 ± 0.89 ^{eg}	−5.1 ± 1.4 ^{adfg}
CSF t-tau, mean ± sd	0 ± 1 ^{cd}	0.726 ± 2.5 ^f	0.791 ± 2.2 ^{cg}	2.39 ± 2.2 ^{dfg}	0 ± 1 ^{bd}	1.06 ± 1.6 ^{bef}	0.0525 ± 1 ^{eg}	2.55 ± 2.6 ^{dfg}
CSF p-tau, mean ± sd	0 ± 1 ^{cd}	0.242 ± 1.5 ^{af}	0.318 ± 1.4 ^{acg}	1.76 ± 1.7 ^{dfg}	0 ± 1 ^{bd}	1.01 ± 1.6a ^{bef}	−0.194 ± 0.69 ^{aeg}	1.91 ± 1.6 ^{dfg}

Memory scores in EMIF are age-, sex- and education-adjusted Z-scores based on published test reference values or local norms; memory scores in ADNI were age-, sex- and education-adjusted Z-scores based on memory scores within the ADNI cohort, using all available memory scores in this population for CN individuals. Levels of amyloid, t-tau and p-tau are Z-scores relative to the CN A- group

CN cognitively normal, MCI mild cognitive impairment

a-g, *p*-value < 0.05 for a: comparing same cognitive and amyloid group between EMIF-AD MBD and ADNI, b: comparing CN A+ and CN A- in same dataset, c: comparing MCI A- and CN A- in same dataset, d: comparing MCI A+ and CN A- in same dataset, e: comparing MCI A- and CN A+ in same dataset, f: comparing MCI A+ and CN A+ in same dataset, g: comparing MCI A+ and MCI A- in same dataset

increased in EMIF-AD, but none of these proteins were measured in ADNI. Therefore, we repeated the analysis in the 48 CSF panel [11] (supplementary Table 2). We found that the 10 synaptic proteins, including VGF and YWHAZ, measured in this panel now showed consisted findings with EMIF-AD MBD. Relative to controls, individuals with preclinical AD and prodromal AD showed lower levels of VGF, NPTX2 and SCG2 which was in concordance with the protein levels observed in EMIF-AD. In addition, YWHAZ was increased in prodromal AD compared to controls and preclinical AD which was also in agreement to the YWHAZ levels observed in EMIF-AD.

Associations of synaptic protein levels with memory scores

Here, we will summarise the results separately for immediate and delayed recall on memory tests, and separately report associations across diagnostic groups vs. associations that depended on diagnostic group (for full results,

see Supplementary Table 1, and kwesenhagen.shinyapps.io/Synaptic_protein_associations_with_memory). First, a lower score on immediate recall was associated with lower concentrations of synaptic proteins in EMIF-AD (49 proteins) and ADNI (6 proteins) (Fig. 3). Of these associations, 1 protein (CDH13) between cohorts. A lower memory score was associated with higher levels of coagulation related proteins in EMIF-AD and with NEFL in ADNI.

In EMIF-AD MBD, 30 synaptic proteins showed an interaction with diagnostic group on immediate recall scores. In prodromal AD, lower levels of 27 proteins were associated with lower immediate recall scores (Fig. 4), while higher levels of only (APOA4) was associated with worse immediate recall. Only 1 to 2 associations were found with immediate memory in controls, preclinical AD and non-AD MCI (Fig. 4, Supplementary Table 1).

In ADNI, 8 proteins showed an interaction with diagnostic group on immediate recall. Lower levels of 7 of

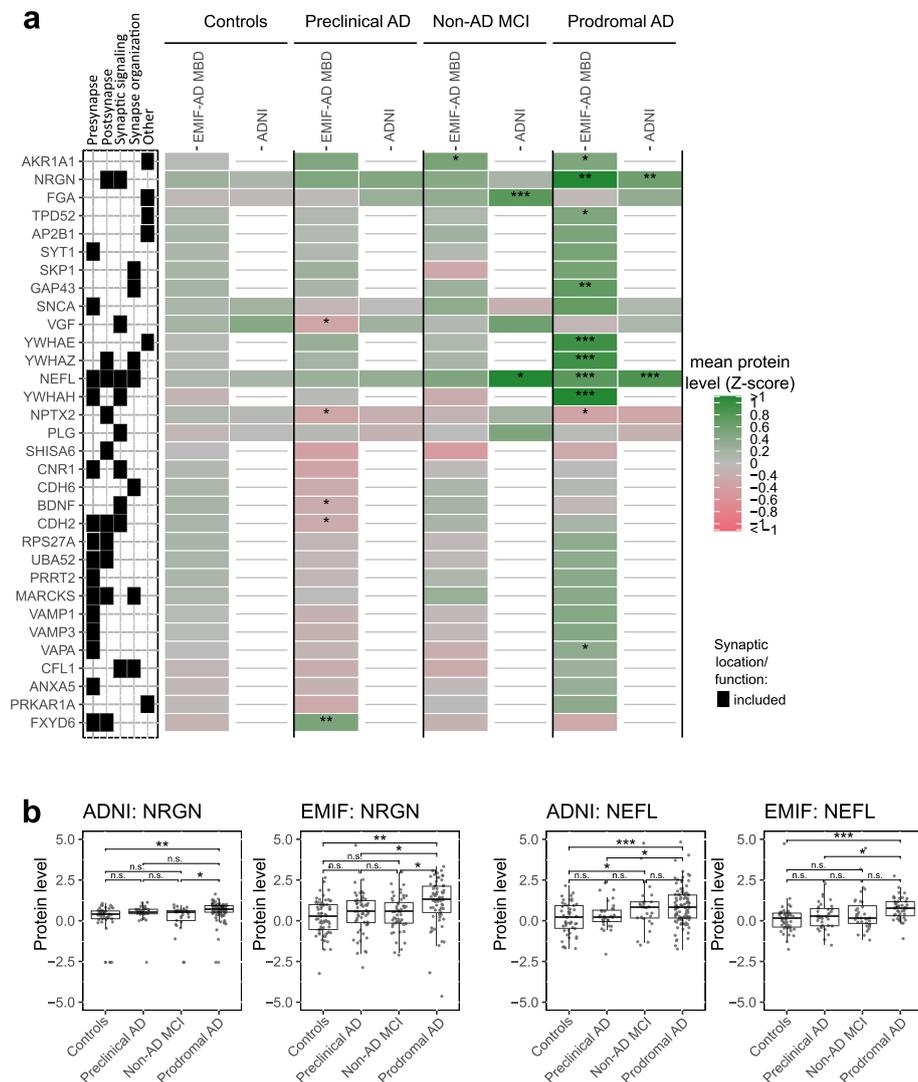


Fig. 2 Differences in synaptic proteins depending on diagnostic group. **a** Mean protein levels are shown for top proteins depending on amyloid and cognitive subgroups, **b** boxplots of protein levels between diagnostic groups for neurogranin (NRGN) and neurofilament light (NEFL). The box of the boxplot indicates 25th percentile, median and 75th percentile, whiskers indicate 1.5 × interquartile range. See Supplementary Table 2 for mean differences between groups and full synaptic annotation for all proteins. Synaptic category ‘Other’ refers to other synaptic locations and functions as detailed in Fig. 1c. *, ** and *** indicate significant difference between diagnostic group and controls (**a**), or between indicated diagnostic groups (**b**): *, *p*-value < 0.05, **, *p*-value < 0.01, ***, *p*-value < 0.001, n.s.: not significant

these proteins (NECTIN, NCSTN, NCSTN, NPTXR, CNTN2, NRXN1, APP) were associated with worse immediate recall in non-AD MCI (NECTIN1, NCSTN, NCSTN, NPTXR, NRXN1 and APP; Fig. 4).

For delayed recall, lower levels of 3 proteins in EMIF-AD MBD and higher levels of 1 protein in ADNI were associated with worse delayed recall independent of diagnostic group (Fig. 3, Supplementary Table 1). For 22 proteins in EMIF-AD MBD, the association between synapse protein level and delayed recall differed between diagnostic groups. In controls, higher levels of GPC4 and

ADAM10 were associated with lower recall scores. In preclinical AD, lower levels of 7 proteins were associated with lower delayed recall scores. In prodromal AD, lower levels of 3 proteins were associated with lower delayed recall scores (supplementary Table 1).

For 11 proteins in ADNI, the association between synapse protein level and delayed recall differed between diagnostic groups. In preclinical AD, lower levels of CLSTN1, CLSTN3, SCG2, CADM3 and NRCAM were associated with lower delayed recall scores (Fig. 4). In non-AD MCI and prodromal AD, lower levels of 2 to 3

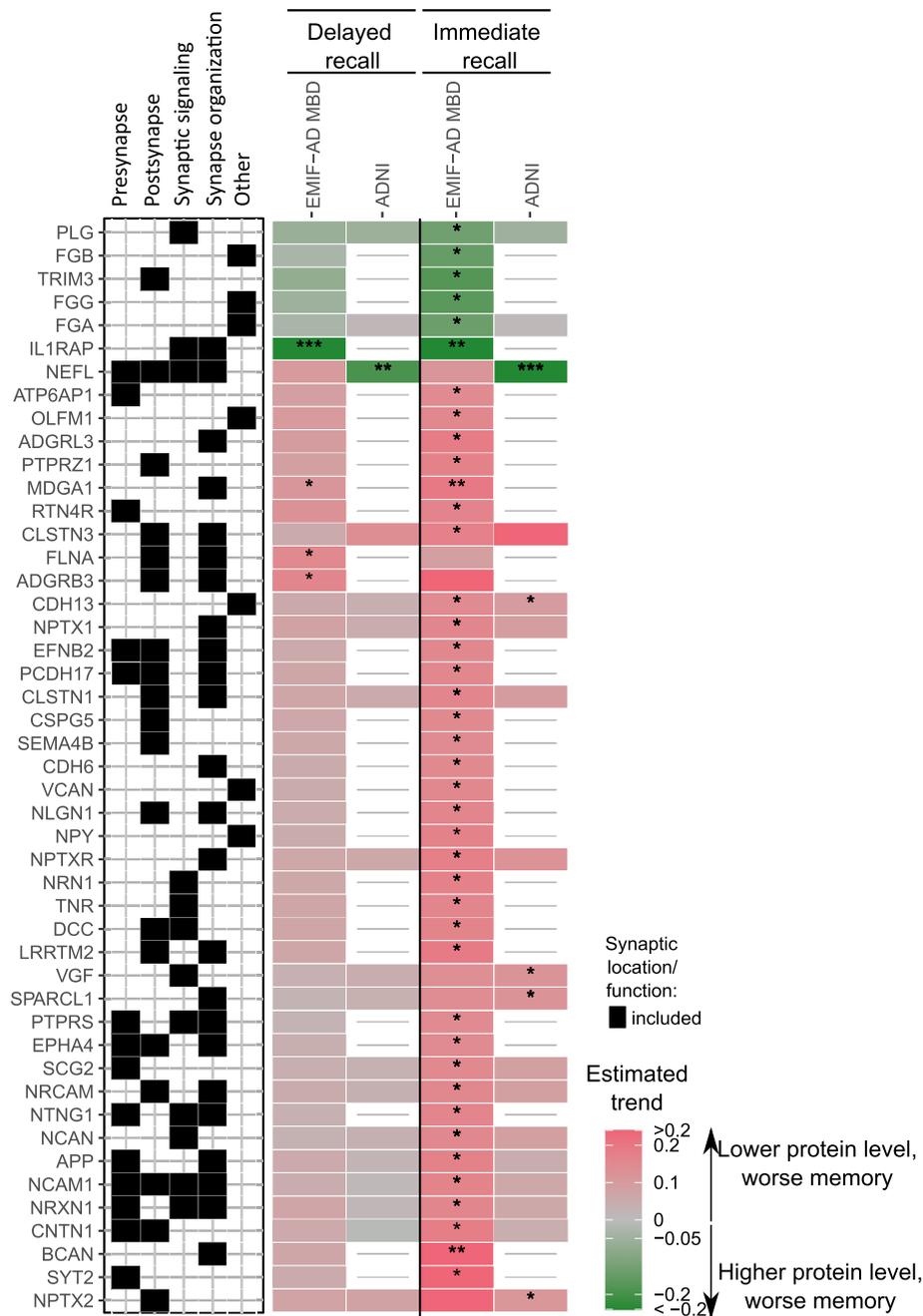


Fig. 3 Synaptic proteins associated with memory independent of diagnostic group. Top proteins associated with memory across all diagnostic groups are shown. Associations were considered significant when proteins did not show an interaction with diagnostic group (interaction p -value ≥ 0.1) and had an association with memory function (*, p -value < 0.05 ; **, p -value < 0.01), ***, p -value < 0.001). Associations of all analysed synaptic proteins are provided in Supplementary Table 1

proteins were associated with worse memory (Fig. 4, Supplementary Fig. 2). Supplementary Table 3 summarizes the findings of protein levels in prodromal AD and non-AD MCI and their associations with memory in both cohorts.

We finally performed enrichment analyses on proteins of which the association with memory scores differed between diagnostic groups. Synaptic proteins that were associated with immediate recall in non-AD MCI showed enrichment for synapse organization, and

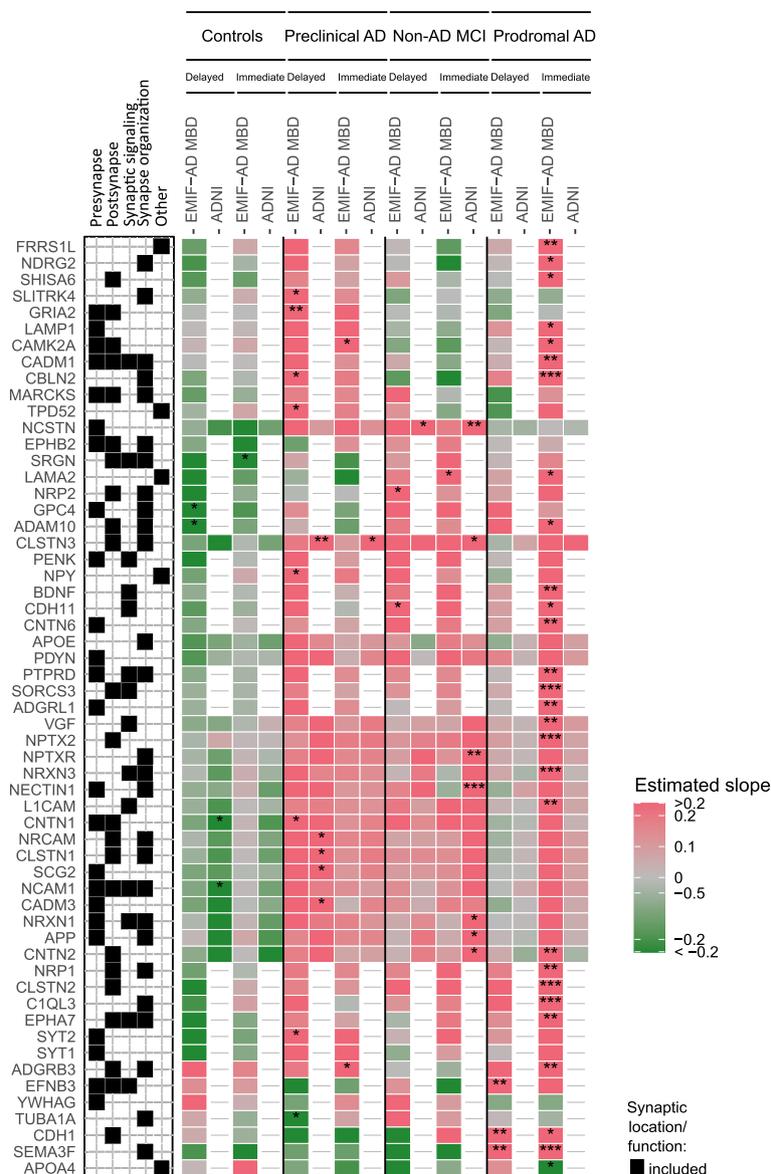


Fig. 4 Memory-associated synaptic proteins stratified for amyloid and cognitive status. Associations between synaptic protein levels and delayed and immediate recall on word learning tests are shown stratified for diagnostic groups based on amyloid and cognitive status in EMIF-AD MBD and ADNI. Proteins which were related to memory function depending on diagnostic group in at least one cohort are shown. Associations were considered significant when proteins showed an interaction with diagnostic group on memory scores (p -value < 0.1) and showed an effect on memory function in diagnostic group-stratified analyses (p -value < 0.05). *, **, ***: significant effect of protein level on memory function in diagnostic group-stratified analyses (*, p -value < 0.05 , **, p -value < 0.01 ; ***, p -value < 0.001). Synaptic category 'Other' refers to other synaptic locations and functions as detailed in Fig. 1c. Associations of all analysed synaptic proteins are provided in Supplementary Table 1

proteins related with immediate recall in prodromal AD showed enrichment for synapse organization and trans-synaptic signalling (Supplementary Fig. 2). Synaptic proteins related with delayed recall in preclinical AD showed enrichment for synapse organization and

presynaptic functions (Supplementary Fig. 3). Figure 5 summarizes the associations of synaptic proteins with memory scores through composite scores of synaptic proteins associated with delayed and immediate recall in EMIF-AD MBD and ADNI.

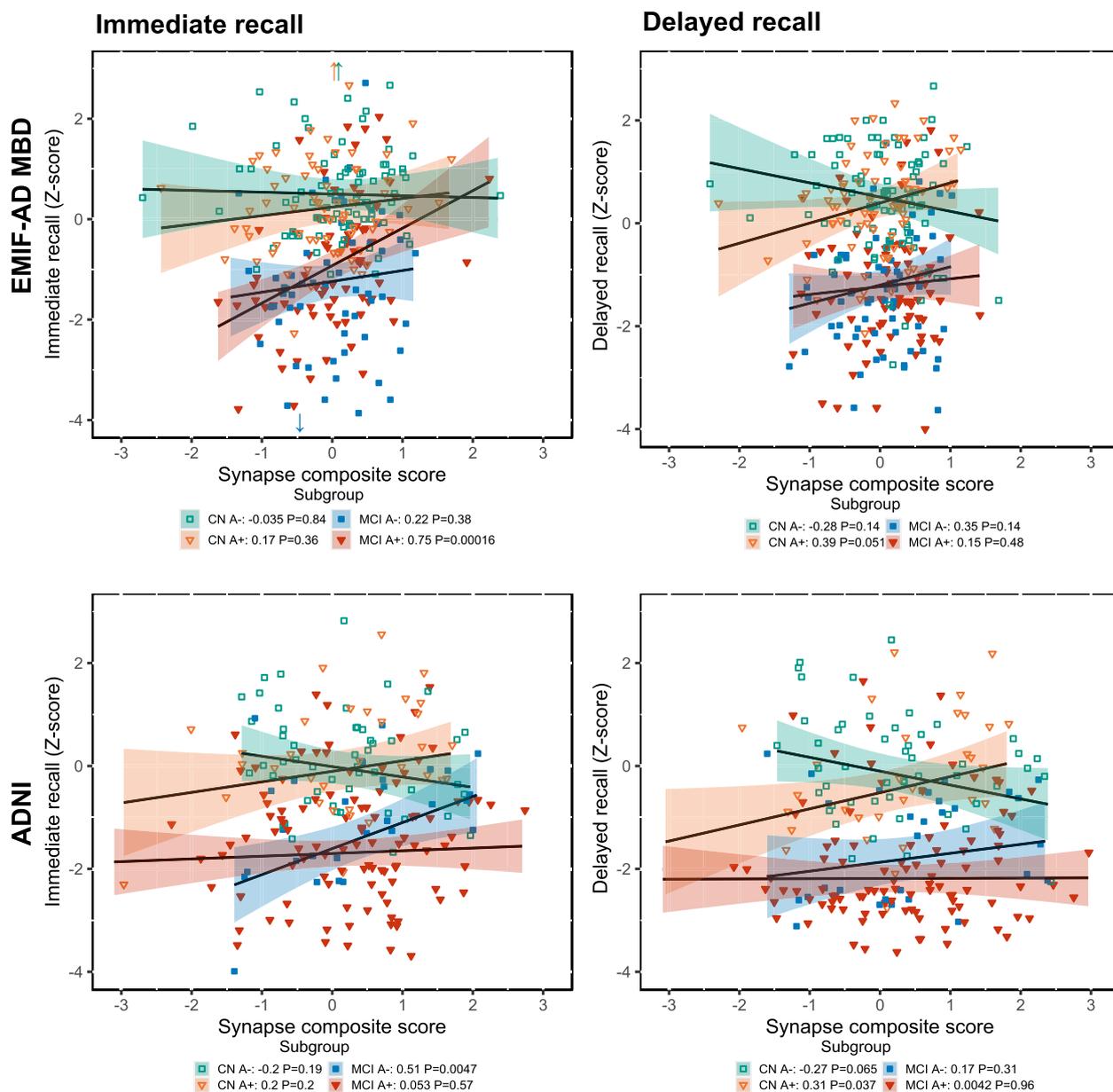


Fig. 5 Associations of synaptic composite scores with immediate and delayed memory scores. Synaptic composites contain proteins associated with delayed recall (left) or immediate recall (right). CN A-, controls; CN A+, preclinical AD, MCI A-, non-AD MCI, MCI A+, prodromal AD

Discussion

Our main finding is that lower synaptic protein levels in CSF were associated with more impaired memory function in predementia AD. Therefore, treatment targeting synapses may be beneficial in early phases of AD.

Reduced synaptic protein levels associated with worse memory functioning in preclinical AD

We found that the concentrations of five synaptic proteins (VGF, CDH2, NPTX2, BDNF and FXD6) were reduced in preclinical AD relative to controls in

EMIF-AD MBD. Reduced synaptic protein levels in CSF have been reported before in preclinical AD [24]. We extended previous studies by showing that lower delayed recall scores were associated with decreased synaptic protein levels in CSF in preclinical AD. These associations may reflect synapse loss [27, 28, 35–40]. The deterioration of synapses could in turn result in memory impairment [2, 8, 47]. Alternatively, this association can be bidirectionally interpreted, with higher levels of these proteins correlating with improved delayed recall scores. As the associated proteins did not show actual decreased levels compared to controls, it remains a possibility that higher protein levels could compensate for neuronal loss.

Associations of synaptic proteins with immediate and delayed recall in prodromal AD

We found higher levels of NRG1 in prodromal AD relative to controls which is in line with previous studies that showed that higher NRG1 levels are associated with advanced disease stage and worse cognition [6, 19, 21]. In our study, however, there was only a slight trend towards a relationship between increased NRG1 levels and memory function, which could imply that NRG1 also relates to other aspects such as disease severity. Previous studies have shown an association of lower levels of NPTX2 and VGF with worse cognition in non-demented individuals with AD and conversion to dementia [20, 22, 23]. This is in line with our results as we have found that decreases in NPTX2 and VGF levels were also related to worse cognition. These results suggest that these proteins may play a role in synaptic disturbance in AD which leads to cognitive impairment and reinforces the prognostic value of these proteins. Overall, the associations of synaptic proteins with immediate and delayed recall were similar, but substantially more associations with immediate recall than delayed recall were found in prodromal AD subjects suggesting that immediate recall may be more sensitive in early AD.

Associations of synaptic proteins with immediate and delayed recall in non-AD MCI

In non-AD MCI, 11 proteins were associated with memory. The majority of memory-associated proteins in non-AD MCI (7 proteins) were specifically related to memory in non-AD MCI. Therefore, these may reflect synaptic dysfunction and memory decline caused by different disorders. For example, for diseases such as Parkinson's disease (PD) and dementia with Lewy bodies (DLB) that can underlie non-AD MCI [7, 26], aggregated alpha synuclein could induce synaptic disturbances [10, 46].

Associations of synaptic proteins with delayed recall in controls

Rather surprisingly, in controls a higher concentration of 5 synaptic proteins were associated with worse delayed recall (Fig. 4). These proteins were not associated with memory in any of the other diagnostic groups. Some controls in EMIF-AD MBD had increased tau levels, which may have contributed to low memory scores in these individuals, as tau can impair synaptic plasticity and cause synaptic damage (Hu et al., n.d.; [17, 30, 33]). However, the proteins that did associate with memory in the other diagnostic groups showed associations of lower levels with lower memory scores. This suggests that the synaptic processes leading to impaired memory may differ in controls, non-AD MCI and Alzheimer's disease. Moreover, we previously showed in cognitively normal individuals with normal AD biomarkers that those with higher synaptic proteins levels had a higher risk for AD pathology at follow-up such that the increase may indicate early AD [43].

Difference in associations between ADNI and EMIF-AD MBD

In general, CSF proteins showed similar associations with memory scores when comparing the EMIF-AD MBD and ADNI datasets. An exception was prodromal AD, in which individuals from the EMIF-AD MBD cohort showed many associations of lower synaptic protein levels and lower immediate recall, which were not observed in ADNI. Potentially, the lack of reproducibility between the cohorts is due to the different word learning tests used in EMIF-AD MBD and ADNI. Prodromal AD individuals in ADNI also had on average lower delayed and immediate recall scores compared to EMIF-AD MBD (Table 1), likely because in ADNI impairment on a memory test was an inclusion criterion for the prodromal AD group, which was not the case in EMIF-AD MBD. The difference in disease severity in prodromal AD and the number and difference in measurement of proteins between both cohorts may also explain the lack of reproducibility. The latter is reinforced by the observation that the same proteins in the ADNI Caprion/RBM dataset showed a different association with cognitive function compared to the same proteins measured in the CSF 48 panel in ADNI.

Memory associations of synaptic subcomponents and functions

In general, we observed that synaptic components (the pre- and post-synapse) and functions (synaptic signaling) showed similar memory associations. This suggests that generalized synapse loss underlies the associations of

the observed synaptic proteins with memory. However, in preclinical AD, we observed enrichment of synapse organization among proteins that were associated with delayed recall in both cohorts. This could imply synapse (re-)organization might be a process involved in memory functioning in very early AD.

Strengths and limitations

A limitation of this study is that cohorts showed different demographics (i.e. ADNI had an older population and lower memory scores), used different memory tests, procedures to normalize memory scores, different criteria for the diagnostic groups, and different proteomic platforms. Nonetheless, we found similar associations of synaptic protein levels with delayed recall in both cohorts and the validation CSF panel, which indicated these results are robust for cohort-dependent effects. For associations of the proteins with immediate recall in prodromal AD and non-AD MCI, we observed more heterogeneity between cohorts, perhaps reflecting that individuals with MCI form a heterogeneous group. This stresses the importance of new, larger studies in individuals with prodromal AD.

Our data included only a part of the synaptic proteome (16.5%), so future studies should target a larger part of the synaptic proteome to investigate the role of synapses in memory functioning in more detail. Furthermore, while CSF proteomics allow simultaneous measurement of many proteins, it is not possible to determine whether alterations in concentrations are specific to particular anatomical brain structures. As such, future studies combining CSF proteomics and synapse PET would provide great anatomical detail of alterations in synaptic density. Lastly, the associations with memory performance we reported were not corrected for multiple testing. Instead, we tested associations with memory performance in two independent datasets which improves the robustness of the results. Strengths of our study are a the relatively large sample size from 2 independent studies and our analysis of synaptic changes in the very early stage of the disease that has received relatively little attention. Another strength is the publication of the memory associations of the analyzed synaptic proteins in an interactive online database (available at kwesenhagen.shinyapps.io/Synaptic_protein_associations_with_memory).

Conclusions

CSF levels of synaptic proteins are decreased in early AD and were associated with memory loss. This indicates that the synapse may be an attractive target for therapeutic modulation in early AD. Further studies should therefore aim to study longitudinal relationships across different stages of AD.

Abbreviations

AD	Alzheimer's disease
ADAM10	ADAM 10
ADGRB3	adhesion G protein-coupled receptor B3
ADNI	Alzheimer's Disease Neuroimaging Initiative
AKR1A1	Aldo-keto reductase family 1 member A1
ANOVA	Analysis of variance
APOE	Apolipoprotein E
APP	amyloid precursor protein
AVLT	auditory verbal learning test
BDNF	brain-derived neurotrophic factor
CADM3	cell adhesion molecule 3
CDH1	cadherin-1
CDH2	cadherin-2
CDH11	cadherin-11
CDH13	cadherin-13
CDR	Clinical Dementia Rating
CERAD	Consortium to Establish a Registry for Alzheimer's Disease wordlist
CLSTN1	calsyntenin-1
CLSTN3	calsyntenin-3
CN	cognitively normal
CNTN1	contactin-1
CNTN2	contactin-2
CSF	cerebrospinal fluid
DLB	dementia with Lewy bodies
EFNB3	ephrin-B3
EMIF-AD MBD	European Medical Information Network Alzheimer's disease multi-modal biomarker discovery study
FCSRT	Free and Cued Selective Reminding Test
FDR	false discovery rate
FGA	fibrinogen alpha chain
FGB	fibrinogen beta chain
FGG	fibrinogen gamma chain
FLNA	filamin-A
FXYD6	FXYD domain containing ion transport regulator 6
GAP43	neuromodulin (also known as growth-associated protein 43)
GPC4	glypican-4
GRIA4	glutamate receptor 4
IL1RAP	IL-1 receptor accessory protein
LAMA2	laminin subunit alpha-2
LC/MS-MRM	liquid chromatography-mass spectrometry with multiple reaction monitoring
LRRTM2	leucine-rich repeat transmembrane neuronal protein 2
MCI	mild cognitive impairment
MDGA1	MAM domain-containing protein 3
MMSE	Mini-Mental State Examination
MRI	magnetic resonance imaging
NCSTN	nicastrin
NECTIN1	nectin-1
NEFL	neurofilament light
NLGN2	neuroligin-2
non-AD MCI	MCI with normal amyloid
NPTX2	neuronal pentraxin-2
NPTXR	neuronal pentraxin receptor
NRCAM	neuronal cell adhesion molecule
NRGN	neurogranin
NRXN1	neurexin-1
NRXN3	neurexin-3
PD	Parkinson's disease
PET	positron emission tomography
PLG	plasminogen
RAVLT	Rey auditory verbal learning test
RBM	Rules-Based Medicine
SCG2	secretogranin-2
SEMA3F	Semaphorin-3F
SNAP25	synaptosomal-associated protein 25
SPARCL1	SPARC-like protein 1
SynGO	Synaptic Gene Ontologies
TMT	tandem mass tag
TPD52	tumor protein D52

TRIM3	tripartite motif-containing protein 3
VAPA	VAMP-associated protein A
VGF	neurosecretory protein VGF
YWHAH	14-3-3 epsilon
YWHAH	14-3-3 eta
YWHAZ	14-3-3 zeta

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13195-025-01703-z>.

- Supplementary Material 1
- Supplementary Material 2
- Supplementary Material 3
- Supplementary Material 4
- Supplementary Material 5
- Supplementary Material 6
- Supplementary Material 7

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Authors' contributions

KW was responsible for data analysis. Both KW and DL contributed equally to writing this article. EMIF-AD: JG, HZ. Supervision: PJV, BMT. JG, HZ, PJV, BMT, JT, IB, SV, PML, MT, JP, GP, MT, RV, YFL, FV, SL, JS, VD, KB, PS, AS, LB and CT read and approved the final manuscript.

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Data availability

EMIF-AD MBD data can be requested by contacting P.J. Visser and Stephanie Vos. ADNI data is publicly available and can be accessed via adni.loni.usc.edu. ADNI data can be downloaded from adni.loni.usc.edu. The raw proteomic data from EMIF-AD MBD has been submitted to the ProteomeXchange Consortium through the PRIDE partner repository, under the dataset identifier <https://doi.org/10.6019/PXD019910>. Requests for access to other EMIF-AD MBD data should be directed to the authors. Data sharing restrictions may apply due to consent agreements from participants in each cohort and European GDPR regulations, which limit data sharing with several non-European countries. Statistical data are provided in the supplementary information files.

Declarations

Ethics approval and consent to participate

For both ADNI and EMIF-AD MBD, Local institutional review boards approved the procedures for this study and written informed consent was obtained in accordance with the Declaration of Helsinki. Supplementary Table 4 contains a full overview of Ethical Committee/IRB names of each center.

Consent for publication

Not applicable.

Competing interests

KB has served as a consultant, at advisory boards, or at data monitoring committees for Abcam, Axon, BioArctic, Biogen, JOMDD/Shimadzu, Julius Clinical, Lilly, MagQu, Novartis, Pharmatrophix, Prothena, Roche Diagnostics, and Siemens Healthineers, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program, outside the work presented in this paper. HZ has served at scientific advisory boards and/or as a consultant for Abbvie, Alector, Annexon, Artery Therapeutics, AZTherapies, CogRx, Denali, Eisai, Nervgen, Pinteon Therapeutics, Red Abbey Labs, Passage Bio, Roche, Samumed, Siemens Healthineers, Triplet Therapeutics, and Wave, has given lectures in symposia sponsored by Cellectricon, Fujirebio, Alzecure, Biogen, and Roche, and is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program. PS has acquired grants for the institution from GE Healthcare and Piramal and received consultancy/speaker fees paid to the institution from Novartis, Probiobdrug, Biogen, Roche, and EIP Pharma, LLC in the past 2 years. CT received grants from the European Commission, the Dutch Research Council (ZonMW), Association of Frontotemporal Dementia/Alzheimer's Drug Discovery Foundation, The Weston Brain Institute, Alzheimer Netherlands. Prof. dr. Teunissen has functioned in advisory boards of Roche, received non-financial support in the form of research consumables from ADXNeurosciences and Euroimmun, performed contract research or received grants from Probiobdrug, Biogen, Esai, Toyama, Janssen Prevention Center, Boehringer, AxonNeurosciences, EIP farma, PeopleBio, Roche. The other authors report no conflict of interest.

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References

- ADNI General Procedures Manual [WWW Document], 2006. URL http://adni.loni.usc.edu/wp-content/uploads/2010/09/ADNI_GeneralProceduresManual.pdf. Accessed 13 Aug 2020.
- Arendt T. Synaptic degeneration in Alzheimer's disease. *Acta Neuropathol.* 2009;118:167–79. <https://doi.org/10.1007/s00401-009-0536-x>.
- Bath TS, Francavilla C, Olsen JV. Off-line high-pH reversed-phase fractionation for in-depth phosphoproteomics. *J Proteome Res.* 2014;13:6176–86. <https://doi.org/10.1021/pr500893m>.
- Bilgel M, An Y, Lang A, Prince J, Ferrucci L, Jedyak B, Resnick SM. Trajectories of Alzheimer disease-related cognitive measures in a longitudinal sample. *Alzheimer's and Dementia.* 2014;10:735–742.e4. <https://doi.org/10.1016/j.jalz.2014.04.520>.
- Bos I, Vos S, Vandenberghe R, Scheltens P, Engelborghs S, Frisoni G, Molinuevo JL, Wallin A, Lleó A, Popp J, Martinez-Lage P, Baird A, Dobson R, Legido-Quigley C, Sleegers K, Van Broeckhoven C, Bertram L, ten Kate M, Barkhof F, Zetterberg H, Lovestone S, Streffer J, Visser PJ. The EMIF-AD Multimodal Biomarker Discovery study: design, methods and cohort characteristics. *Alzheimers Res Ther.* 2018;10:64. <https://doi.org/10.1186/s13195-018-0396-5>.
- Casaleto KB, Elahi FM, Bettcher BM, Neuhaus J, Bendlin BB, Asthana S, Johnson SC, Yaffe K, Carlsson C, Blennow K, Zetterberg H, Kramer JH. Neurogranin, a synaptic protein, is associated with memory independent of Alzheimer biomarkers. *Neurology.* 2017;89:1782–8. <https://doi.org/10.1212/WNL.0000000000004569>.
- Ciafone J, Little B, Thomas AJ, Gallagher P. The Neuropsychological Profile of Mild Cognitive Impairment in Lewy Body Dementias. *J Int Neuropsychol Soc.* 2020;26:210–25. <https://doi.org/10.1017/S1355617719001103>.
- Colom-Cadena M, Spiers-Jones T, Zetterberg H, et al. The clinical promise of biomarkers of synapse damage or loss in Alzheimer's disease. *Alz Res Ther.* 2020;12:21. <https://doi.org/10.1186/s13195-020-00588-4>.
- Dulewicz M, Kulczyńska-Przybik A, Mroczko B. Neurogranin and VILIP-1 as Molecular Indicators of Neurodegeneration in Alzheimer's Disease: A Systematic Review and Meta-Analysis. *Int J Mol Sci.* 2020;21:8335. <https://doi.org/10.3390/ijms21218335>.
- Gcwensa NZ, Russell DL, Cowell RM, Volpicelli-Daley LA. Molecular Mechanisms Underlying Synaptic and Axon Degeneration in Parkinson's Disease. *Front Cell Neurosci.* 2021;0:44. <https://doi.org/10.3389/FNCEL.2021.626128>.
- Haque R, Watson CM, Liu J, Carter EK, Duong DM, Lah JJ, Wingo AP, Roberts BR, Johnson ECB, Saykin AJ, Shaw LM, Seyfried NT, Wingo TS, Levey AI. A protein panel in cerebrospinal fluid for diagnostic and predictive assessment of Alzheimer's disease. *Science translational medicine.* 2023;15(712):4122. <https://doi.org/10.1126/scitranslmed.adg4122>.
- Headley A, De Leon-Benedetti A, Dong C, Levin B, Loewenstein D, Camargo C, Rundek T, Zetterberg H, Blennow K, Wright CB, Sun X. Neurogranin as a predictor of memory and executive function decline in MCI patients. *Neurology.* 2018;90:e887–95. <https://doi.org/10.1212/WNL.0000000000005057>.
- Jack CR, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, Holtzman DM, Jagust W, Jessen F, Karlawish J, Liu E, Molinuevo JL, Montine T, Phelps C, Rankin KP, Rowe CC, Scheltens P, Siemers E, Snyder HM, Sperling R, Elliott C, Masliah E, Ryan L, Silverberg N. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimer's & Dementia.* 2018;14:535–62. <https://doi.org/10.1016/J.JALZ.2018.02.018>.
- Jansen WJ, Ossenkoppele R, Knol DL, Tijms BM, Scheltens P, Verhey FRJ, Visser PJ, Aarsland D, Alcolea D, Alexander M, Almdahl IS, Arnold SE, Baldeiras I, Barthel H, van Berckel BNM, Bibeau K, Blennow K, Brooks DJ, van Buchem MA, Camus V, Cavedo E, Chen K, Chetelat G, Cohen AD, Drzezga A, Engelborghs S, Fagan AM, Fladby T, Fleisher AS, van der Flier WM, Ford L, Förster S, Fortea J, Foskett N, Frederiksen KS, Freund-Levi Y, Frisoni GB, Froelich L, Gabryelewicz T, Gill KD, Gkatzima O, Gómez-Tortosa E, Gordon MF, Grimmer T, Hampel H, Hausner L, Hellwig S, Herukka S-K, Hildebrandt H, Ishihara L, Ivanoiu A, Jagust WJ, Johannsen P, Kandimalla R, Kapaki E, Klimkiewicz-Mrowiec A, Klunk WE, Köhler S, Koglin N, Kornhuber J, Kramerberger MG, van Laere K, Landau SM, Lee DY, de Leon M, Lisetti V, Lleó A, Madsen K, Maier W, Marcusson J, Mattsson N, de Mendonça A, Meulenbroek O, Meyer PT, Mintun MA, Mok V, Molinuevo JL, Møllergård HM, Morris JC, Mroczko B, van der Mussele S, Na DL, Newberg A, Nordberg A, Nordlund A, Novak GP, Paraskevas GP, Parnetti L, Perera G, Peters O, Popp J, Prabhakar S, Rabinovici GD, Ramakers IHGB, Rami L, Resende de Oliveira C, Rinne JO, Rodrigue KM, Rodrigue-Rodriguez E, Roe CM, Rot U, Rowe CC, Rütger E, Sabri O, Sanchez-Juan P, Santana I, Sarazin M, Schröder J, Schütte C, Seo SW, Soetewey F, Soininen H, Spiru L, Struyfs H, Teunissen CE, Tsolaki M, Vandenberghe R, Verbeek MM, Villemagne VL, Vos SJB, van Waalwijk van Doorn, L.J.C., Waldemar, G., Wallin, A., Wallin, Å.K., Wiltfang, J., Wolk, D.A., Zboch M., Zetterberg, H., Zetterberg, H., Prevalence of Cerebral Amyloid Pathology in Persons Without Dementia. *JAMA.* 2015;313:1924. <https://doi.org/10.1001/jama.2015.4668>.
- Jansen WJ, Ossenkoppele R, Tijms BM, Fagan AM, Hansson O, Klunk WE, van der Flier WM, Villemagne VL, Frisoni GB, Fleisher AS, Lleó A, Mintun MA, Wallin A, Engelborghs S, Na DL, Chetelat G, Molinuevo JL, Landau SM, Mattsson N, Kornhuber J, Sabri O, Rowe CC, Parnetti L, Popp J, Fladby T, Jagust WJ, Aalten P, Lee DY, Vandenberghe R, Resende de Oliveira C, Kapaki E, Froelich L, Ivanoiu A, Gabryelewicz T, Verbeek MM, Sanchez-Juan P, Hildebrandt H, Camus V, Zboch M, Brooks DJ, Drzezga A, Rinne JO, Newberg A, de Mendonça A, Sarazin M, Rabinovici GD, Madsen K, Kramerberger MG, Nordberg A, Mok V, Mroczko B, Wolk DA, Meyer PT, Tsolaki M, Scheltens P, Verhey FRJ, Visser PJ, Aarsland D, Alcolea D, Alexander M, Almdahl IS, Arnold SE, Baldeiras I, Barthel H, van Berckel BNM, Blennow K, van Buchem MA, Cavedo E, Chen K, Chipi E, Cohen AD, Förster S, Fortea J, Frederiksen KS, Freund-Levi Y, Gkatzima O, Gordon MF, Grimmer T, Hampel H, Hausner L, Hellwig S, Herukka S-K, Johannsen P, Klimkiewicz-Mrowiec A, Köhler S, Koglin N, van Laere K, de Leon M, Lisetti V, Maier W, Marcusson J, Meulenbroek O, Møllergård HM, Morris JC, Nordlund A, Novak GP, Paraskevas GP, Perera G, Peters O, Ramakers IHGB, Rami L,

- Rodríguez-Rodríguez E, Roe CM, Rot U, Rütther E, Santana I, Schröder J, Seo SW, Soininen H, Spuru L, Stomrud E, Struyfs H, Teunissen CE, Vos SJB, van Waalwijk van Doorn, L.J.C., Waldemar, G., Wallin, Å.K., Wiltfang, J., Zetterberg, H., Association of Cerebral Amyloid- β Aggregation With Cognitive Functioning in Persons Without Dementia. *JAMA Psychiat*. 2018;75:84. <https://doi.org/10.1001/jamapsychiatry.2017.3391>.
16. Jutten RJ, Sikkes SAM, Amariglio RE, Buckley RF, Properzi MJ, Marshall GA, Rentz DM, Johnson KA, Teunissen CE, van Berckel BNM, van der Flier WM, Scheltens P, Sperling RA, Papp K, v., Identifying Sensitive Measures of Cognitive Decline at Different Clinical Stages of Alzheimer's Disease. *J Int Neuropsychol Soc*. 2021;27:426–38. <https://doi.org/10.1017/S155617720000934>.
 17. Kaniyappan S, Chandupatla RR, Mandelkow E-M, Mandelkow E. Extracellular low-n oligomers of tau cause selective synaptotoxicity without affecting cell viability. *Alzheimer's & Dementia*. 2017;13:1270–91. <https://doi.org/10.1016/j.jalz.2017.04.002>.
 18. Koopmans F, van Nierop P, Andres-Alonso M, Byrnes A, Cijssouw T, Coba MP, Cornelisse LN, Farrell RJ, Goldschmidt HL, Howrigan DP, Hussain NK, Imig C, de Jong APH, Jung H, Kohansalnodehi M, Kramarz B, Lipstein N, Lovering RC, MacGillavry H, Mariano V, Mi H, Ninov M, Osumi-Sutherland D, Pielot R, Smalla KH, Tang H, Tashman K, Toonen RFG, Verpelli C, Reig-Viader R, Watanabe K, van Weering J, Achsel T, Ashrafi G, Asi N, Brown TC, De Camilli P, Feuermann M, Foulger RE, Gaudet P, Joglekar A, Kanelloupolos A, Malenka R, Nicoll RA, Pulido C, de Juan-Sanz J, Sheng M, Südhof TC, Tilgner HU, Bagni C, Bayés À, Biederer T, Brose N, Chua JJE, Dieterich DC, Gundelfinger ED, Hoogenraad C, Huganir RL, Jahn R, Kaeser PS, Kim E, Kreuzt MR, McPherson PS, Neale BM, O'Connor V, Posthuma D, Ryan TA, Sala C, Feng G, Hyman SE, Thomas PD, Smit AB, Verhage M. SynGO: An Evidence-Based, Expert-Curated Knowledge Base for the Synapse. *Neuron*. 2019;103:217–234.e4. <https://doi.org/10.1016/j.neuron.2019.05.002>.
 19. Kvarnberg H, Duits FH, Ingelsson M, Andreasen N, Öhrfelt A, Andersson K, Brinkmalm G, Lannfelt L, Minthon L, Hansson O, Andreasson U, Teunissen CE, Scheltens P, van der Flier WM, Zetterberg H, Portelius E, Blennow K. Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal Alzheimer's disease. *Alzheimer's & Dementia*. 2015;11:1180–90. <https://doi.org/10.1016/j.jalz.2014.10.009>.
 20. Libiger O, Shaw LM, Watson MH, Nairn AC, Umaña KL, Biarnes MC, Canet-Avilés RM, Jack CR, Bretton YA, Cortes L, Chelsky D, Spellman DS, Baker SA, Raghavan N, Potter WZ. Longitudinal CSF proteomics identifies NPTX2 as a prognostic biomarker of Alzheimer's disease. *Alzheimer's and Dementia*. 2021. <https://doi.org/10.1002/alz.12353>.
 21. Liu W, Lin H, He X, Chen L, Dai Y, Jia W, Xue X, Tao J, Chen L. Neurogranin as a cognitive biomarker in cerebrospinal fluid and blood exosomes for Alzheimer's disease and mild cognitive impairment. *Translat Psychiat*. 2020;10:1–9. <https://doi.org/10.1038/s41398-020-0801-2>.
 22. Llano DA, Devanarayan P, Devanarayan V, Alzheimer's Disease Neuroimaging Initiative (ADNI). CSF peptides from VGF and other markers enhance prediction of MCI to AD progression using the ATN framework. *Neurobiol Aging*. 2023;121:15–27. <https://doi.org/10.1016/j.neurobiolaging.2022.07.015>.
 23. Llano DA, Devanarayan P, Devanarayan V, Alzheimer's Disease Neuroimaging Initiative (ADNI). VGF in Cerebrospinal Fluid Combined With Conventional Biomarkers Enhances Prediction of Conversion From MCI to AD. *Alzheimer Dis Assoc Disord*. 2019;33(4):307–14. <https://doi.org/10.1097/WAD.0000000000000328>.
 24. Lleó A, Núñez-Llaves R, Alcolea D, Chiva C, Balateu-Pañós D, Colom-Cadena M, Gomez-Giro G, Muñoz L, Querol-Vilaseca M, Pegueroles J, Rami L, Lladó A, Molinuevo J, Tainta M, Clarimón J, Spires-Jones T, Blesa R, Fortea J, Martínez-Lage P, Sánchez-Valle R, Sábido E, Bayés À, Belbin O. Changes in synaptic proteins precede neurodegeneration markers in preclinical Alzheimer's disease cerebrospinal fluid. *Mole Cell Proteomics*. 2019. mcp.RA118.001290. <https://doi.org/10.1074/mcp.RA118.001290>
 25. Magdalinou NK, Noyce AJ, Pinto R, Lindstrom E, Holmén-Larsson J, Holttá M, Blennow K, Morris HR, Skillbäck T, Warner TT, Lees AJ, Pike I, Ward M, Zetterberg H, Gobom J. Identification of candidate cerebrospinal fluid biomarkers in parkinsonism using quantitative proteomics. *Parkinsonism Relat Disord*. 2017;37:65–71. <https://doi.org/10.1016/j.parkreldis.2017.01.016>.
 26. Martínez-Horta S, Kulisevsky J. Mild cognitive impairment in Parkinson's disease. *J Neural Trans*. 2019;126:7 126, 897–904. <https://doi.org/10.1007/S00702-019-02003-1>.
 27. Masliah E, Mallory M, Alford M, DeTeresa R, Hansen LA, McKeel DW, Morris JC. Altered expression of synaptic proteins occurs early during progression of Alzheimer's disease. *Neurology*. 2001;56:127–9. <https://doi.org/10.1212/wnl.56.1.127>.
 28. Masliah E, Mallory M, Hansen L, Richard DT, Alford M, Terry R. Synaptic and neuritic alterations during the progression of Alzheimer's disease. *Neurosci Lett*. 1994;174:67–72. [https://doi.org/10.1016/0304-3940\(94\)90121-X](https://doi.org/10.1016/0304-3940(94)90121-X).
 29. Nilsson, J., Gobom, J., Sjödin, S., Brinkmalm, G., Ashton, N.J., Svensson, J., Johansson, P., Portelius, E., Zetterberg, H., Blennow, K., Brinkmalm, A., 2021. Cerebrospinal fluid biomarker panel for synaptic dysfunction in Alzheimer's disease. *Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring* 13. <https://doi.org/10.1002/dad2.12179>
 30. Ondrejčák T, Hu N-W, Qi Y, Klyubin I, Corbett GT, Fraser G, Perkinson MS, Walsh DM, Billinton A, Rowan MJ. Soluble tau aggregates inhibit synaptic long-term depression and amyloid β -facilitated LTD in vivo. 2019. <https://doi.org/10.1016/j.nbd.2019.03.022>.
 31. Pereira JB, Janelidze S, Ossenkoppele R, Kvarnberg H, Brinkmalm A, Mattsson-Carlsson N, Stomrud E, Smith R, Zetterberg H, Blennow K, Hansson O. Untangling the association of amyloid- β and tau with synaptic and axonal loss in Alzheimer's disease. *Brain*. 2020. <https://doi.org/10.1093/brain/awaa395>.
 32. Petersen RC. Mild cognitive impairment as a diagnostic entity. *J Intern Med*. 2004;256:183–94. <https://doi.org/10.1111/j.1365-2796.2004.01388.x>.
 33. Piacentini R, Puma DDL, Mainardi M, Lazzarino G, Tavazzi B, Arancio O, Grassi C. Reduced gliotransmitter release from astrocytes mediates tau-induced synaptic dysfunction in cultured hippocampal neurons. *Glia*. 2017;65:1302–16. <https://doi.org/10.1002/GLIA.23163>.
 34. Portelius E, Zetterberg H, Skillbäck T, Törnqvist U, Andreasson U, Trojanowski JQ, Weiner MW, Shaw LM, Mattsson N, Blennow K. Cerebrospinal fluid neurogranin: Relation to cognition and neurodegeneration in Alzheimer's disease. *Brain*. 2015;138:3373–85. <https://doi.org/10.1093/brain/awv267>.
 35. Scheff SW, DeKosky ST, Price DA. Quantitative assessment of cortical synaptic density in Alzheimer's disease. *Neurobiol Aging*. 1990;11:29–37. [https://doi.org/10.1016/0197-4580\(90\)90059-9](https://doi.org/10.1016/0197-4580(90)90059-9).
 36. Scheff SW, Price DA. Synaptic density in the inner molecular layer of the hippocampal dentate gyrus in Alzheimer disease. *J Neuropathol Exp Neurol*. 1998;57:1146–53. <https://doi.org/10.1097/00005072-199812000-00006>.
 37. Scheff SW, Price DA, Schmitt FA, Mufson EJ. Hippocampal synaptic loss in early Alzheimer's disease and mild cognitive impairment. *Neurobiol Aging*. 2006;27:1372–84. <https://doi.org/10.1016/j.neurobiolaging.2005.09.012>.
 38. Scheff SW, Price DA, Schmitt FA, Roberts KN, Ikonovic MD, Mufson EJ. Synapse Stability in the Precuneus Early in the Progression of Alzheimer's Disease. *Journal of Alzheimer's Disease*. 2013;35:599–609. <https://doi.org/10.3233/JAD-122353>.
 39. Scheff SW, Price DA, Schmitt FA, Scheff MA, Mufson EJ. Synaptic loss in the inferior temporal gyrus in mild cognitive impairment and Alzheimer's disease. *Journal of Alzheimer's Disease*. 2011;24:547–57. <https://doi.org/10.3233/JAD-2011-101782>.
 40. Sun N, Chan F-Y, Lu Y-J, Neves MAC, Lui H-K, Wang Y, Chow K-Y, Chan K-F, Yan S-C, Leung Y-C, Abagyan R, Chan T-H, Wong K-Y. Rational Design of Berberine-Based FtsZ Inhibitors with Broad-Spectrum Antibacterial Activity. *PLoS ONE*. 2014;9: e97514. <https://doi.org/10.1371/journal.pone.0097514>.
 41. Tarawneh R, D'Angelo G, Crimmins D, Herries E, Griest T, Fagan AM, Zipfel GJ, Ladenson JH, Morris JC, Holtzman DM. Diagnostic and Prognostic Utility of the Synaptic Marker Neurogranin in Alzheimer Disease. *JAMA Neurol*. 2016;73:561. <https://doi.org/10.1001/jamaneurol.2016.0086>.
 42. Tijms BM. Pathophysiological subtypes of Alzheimer's disease based on cerebrospinal fluid proteomics (in press). *Brain*. 2020. <https://doi.org/10.1093/brain/awaa325>.
 43. Tijms BM, Gobom J, Teunissen CE, Dobricic V, Tsolaki M, Verhey F, Popp J, Martínez-Lage P, Vandenberghe R, Lleó A, Molinuevo JL, Engelborghs S, Freund-Levi Y, Froelich L, Bertram L, Lovestone S, Streffer J, Vos S, Adni B, K., Scheltens, P., Zetterberg, H., Visser, P.J. CSF Proteomic Alzheimer's Disease-Predictive Subtypes in Cognitively Intact Amyloid Negative

- Individuals. *Proteomes*. 2021;9(3):36. <https://doi.org/10.3390/proteomes9030036>.
44. Tijms BM, Willemse EAJ, Zwan MD, Mulder SD, Visser PJ, van Berckel BNM, van der Flier WM, Scheltens P, Teunissen CE. Unbiased approach to counteract upward drift in cerebrospinal fluid amyloid- β 1–42 analysis results. *Clin Chem*. 2018;64:576–85. <https://doi.org/10.1373/clinchem.2017.281055>.
 45. Timmers T, Ossenkuppele R, Verfaillie SCJ, van der Weijden CWJ, Slot RER, Wesselman LMP, Windhorst AD, Wolters EE, Yaqub M, Prins ND, Lammermsma AA, Scheltens P, van der Flier WM, van Berckel BNM. Amyloid PET and cognitive decline in cognitively normal individuals: the SCIENCe project. *Neurobiol Aging*. 2019;79:50–8. <https://doi.org/10.1016/J.NEUROBIOLAGING.2019.02.020>.
 46. Trudler D, Sanz-Blasco S, Eisele YS, Ghatak S, Bodhinathan K, Akhtar MW, Lynch WP, Piña-Crespo JC, Talantova M, Kelly JW, Lipton SA. α -Synuclein Oligomers Induce Glutamate Release from Astrocytes and Excessive Extrasynaptic NMDAR Activity in Neurons, Thus Contributing to Synapse Loss. *J Neurosci*. 2021;41:2264–73. <https://doi.org/10.1523/JNEUROSCI.1871-20.2020>.
 47. Tzioras M, McGeachan RI, Durrant CS, et al. Synaptic degeneration in Alzheimer disease. *Nat Rev Neurol*. 2023;19:19–38. <https://doi.org/10.1038/s41582-022-00749-z>.
 48. Winblad B, Palmer K, Kivipelto M, Jelic V, Fratiglioni L, Wahlund LO, Nordberg A, Bäckman L, Albert M, Almkvist O, Arai H, Basun H, Blennow K, De Leon M, Decarli C, Erkinjuntti T, Giacobini E, Graff C, Hardy J, Jack C, Jorm A, Ritchie K, Van Duijn C, Visser P, Petersen RC, 2004. Mild cognitive impairment - Beyond controversies, towards a consensus: Report of the International Working Group on Mild Cognitive Impairment, in: *Journal of Internal Medicine*. *J Intern Med*, pp. 240–246. <https://doi.org/10.1111/j.1365-2796.2004.01380.x>.

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